

Standardized, high-throughput platform for automated, rapid, and extensive plasma proteome characterization

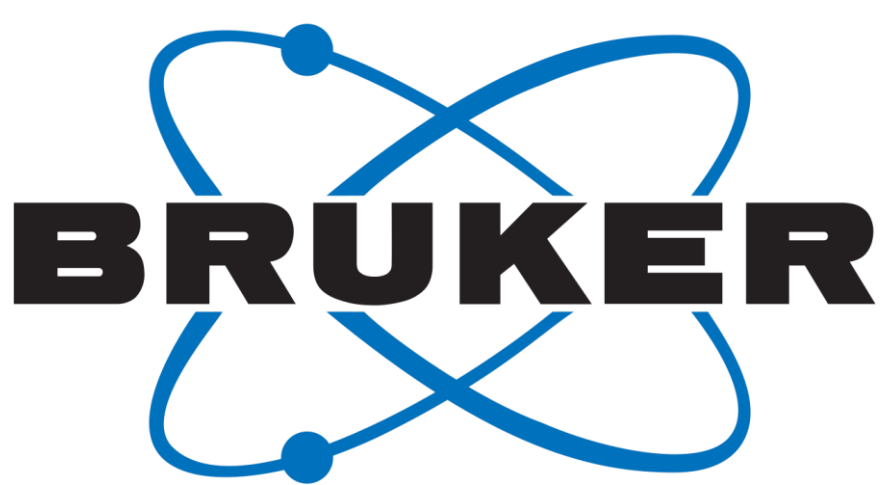


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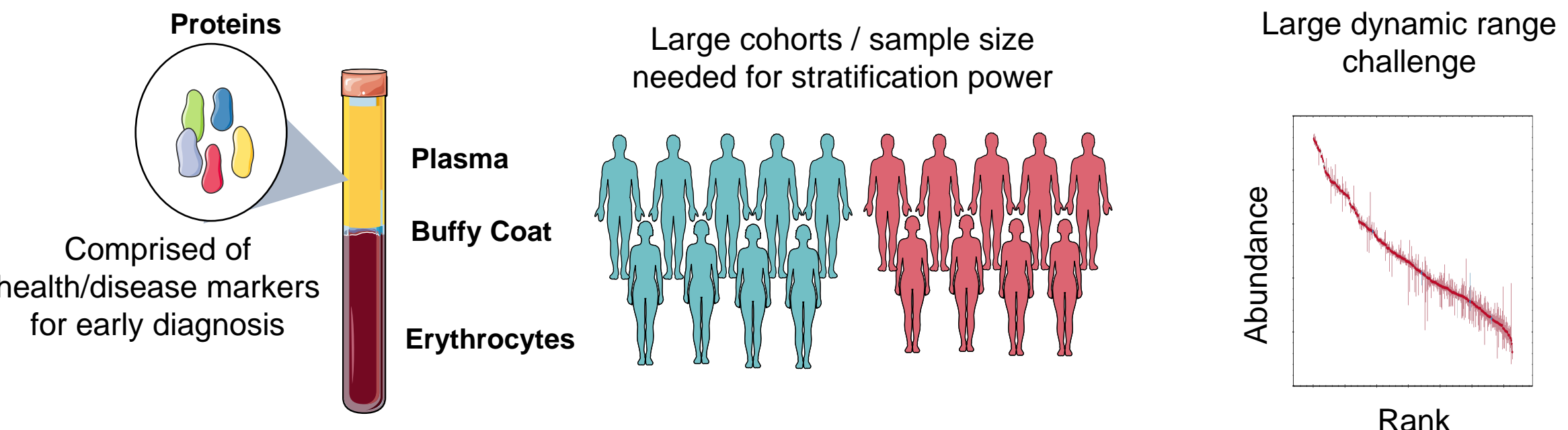
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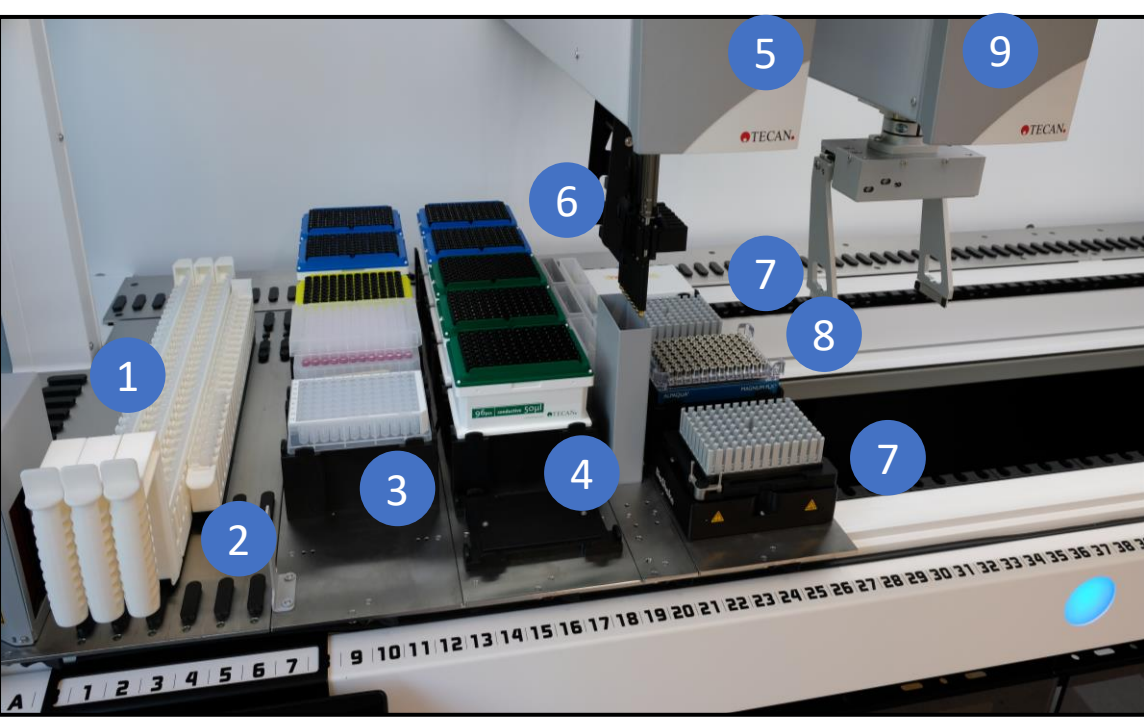
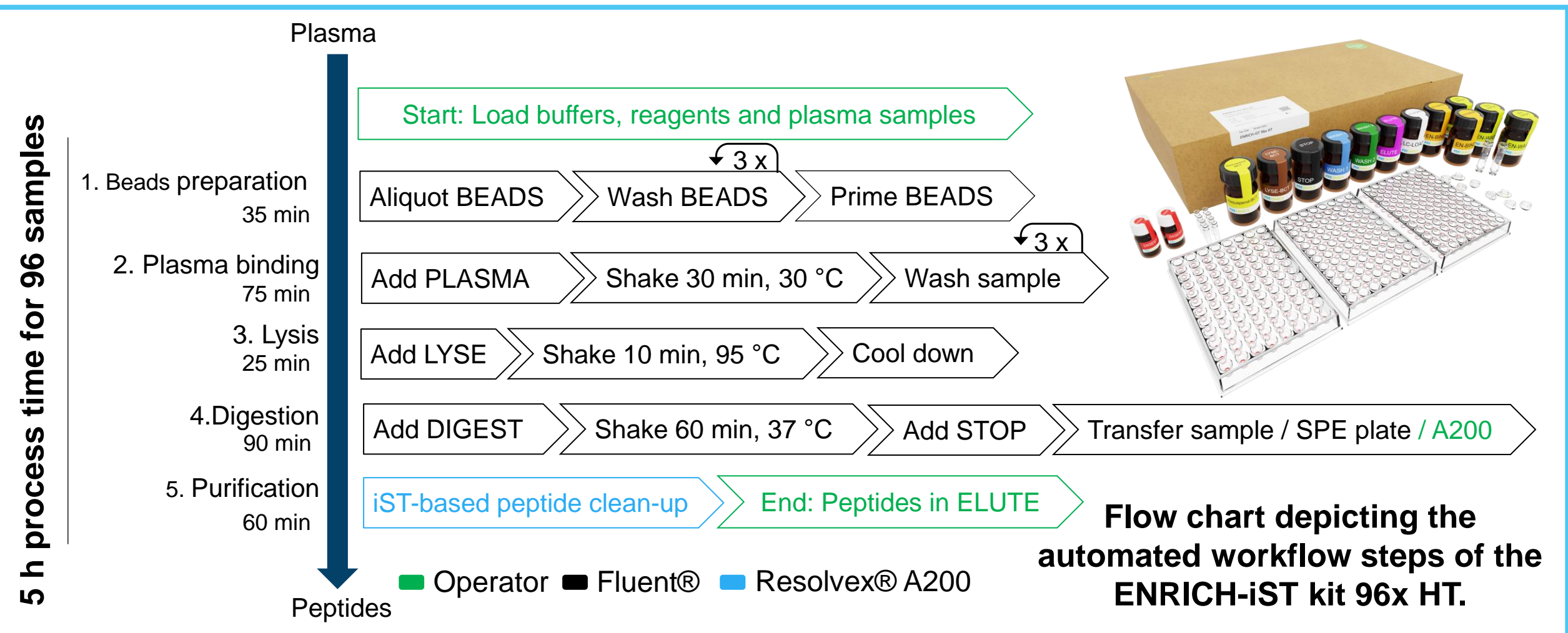
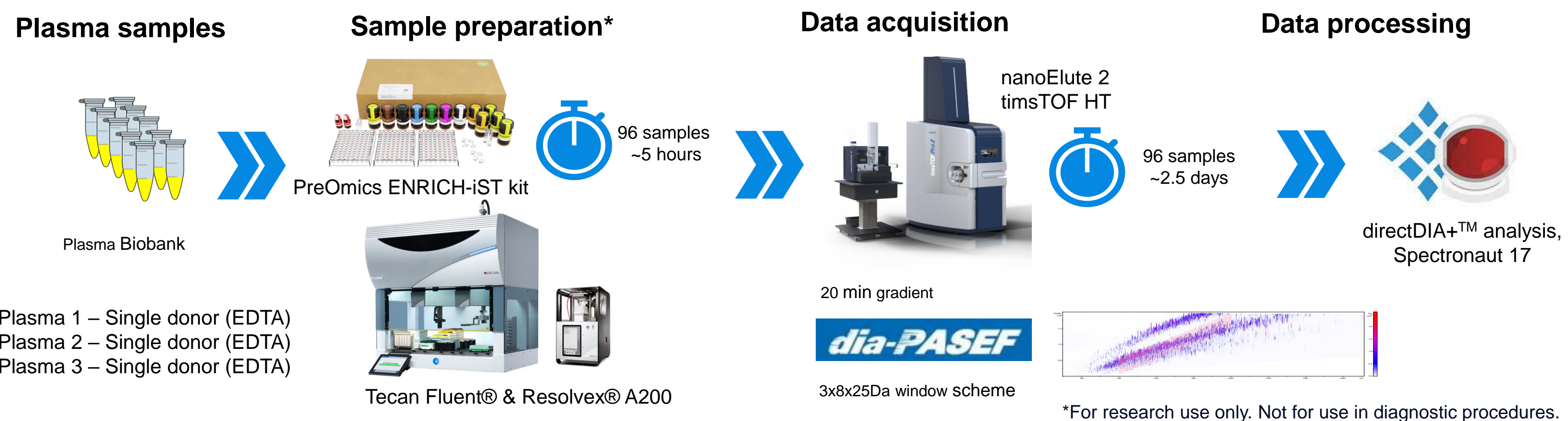
*These authors contributed equally to the work, + Presenting author



Promises & Challenges of Plasma Proteomics

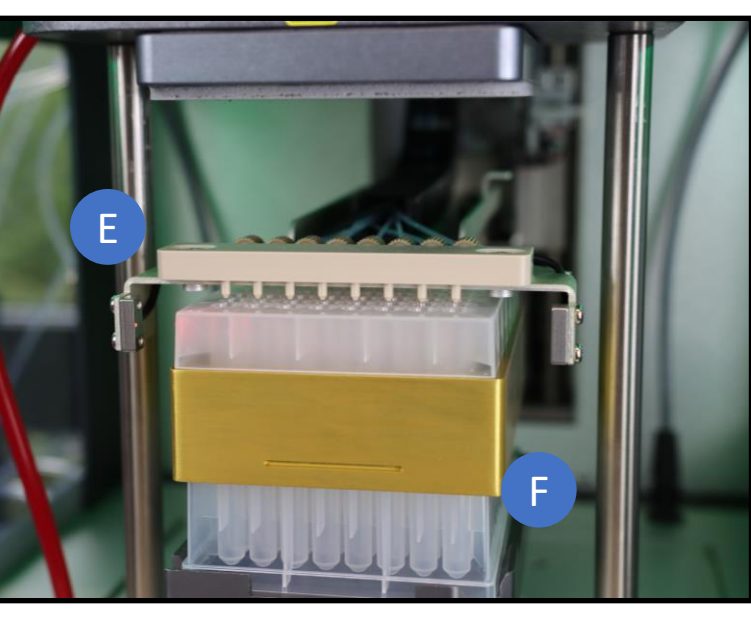
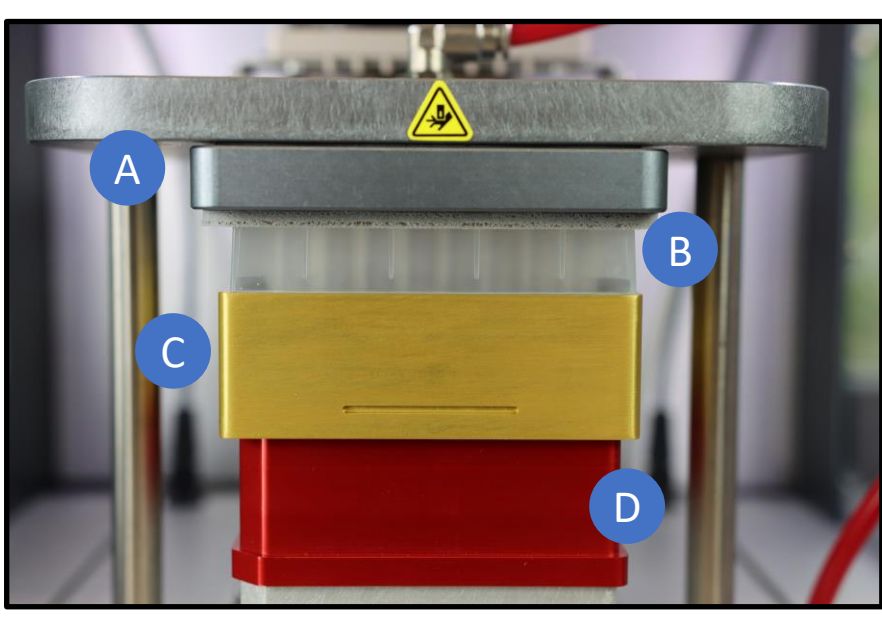


Simplified and scalable plasma proteomics including an in-depth analysis is required to meet challenges of pre-clinical research.



1. Fluent ID™ with 1.5 ml Eppendorf tube runners
2. BEADS & DIGEST storage in 2 ml Sarstedt tubes
3. Assay Plate & 96 well SPE plate
4. 1000 µl, 200 µl, and 50 µl Disposable Tips
5. Flexible Channel Arm™ (AirFCA)
6. 25 ml & 100 ml troughs for WASTE & Buffer storage
7. Bioshake, 37 °C, 95 °C (1200 rpm, 3 mm orbital)
8. Ring-Magnet (Alpaqua FLX)
9. Robotic Gripper Arm (RGA)

Fluent® workdeck layout. 8-channel pipetting (AirFCA) and automated plate transfer (RGA). Liquid level detection facilitates process supervision. On-deck heating/shaking for lysis & tryptic digest. Full reagent & buffer on-deck storage. Starting from Eppendorf tube stored samples, 2 - 96 samples can be prepared fully automated including magnetic bead capture, wash and elution without user intervention.

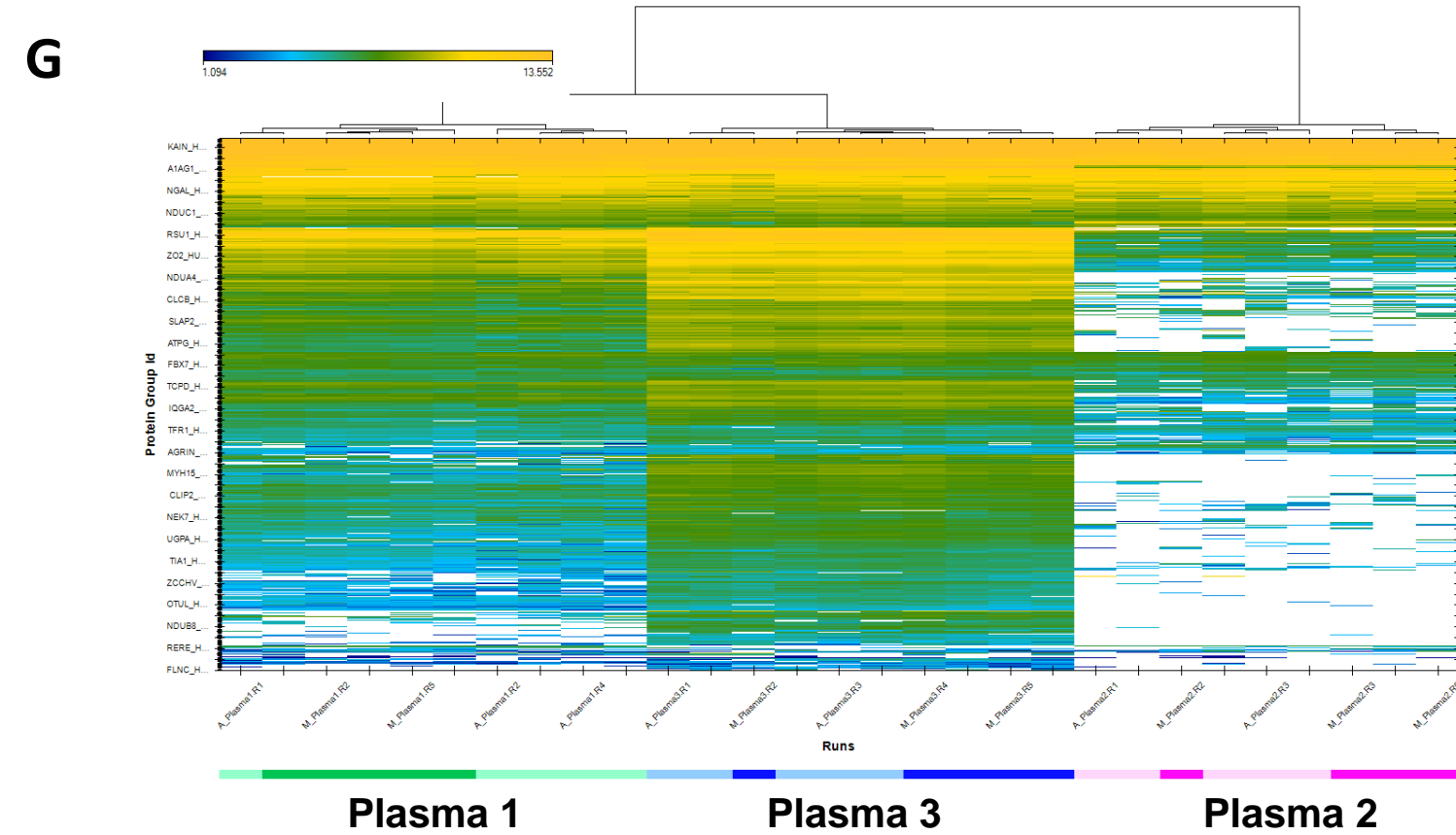
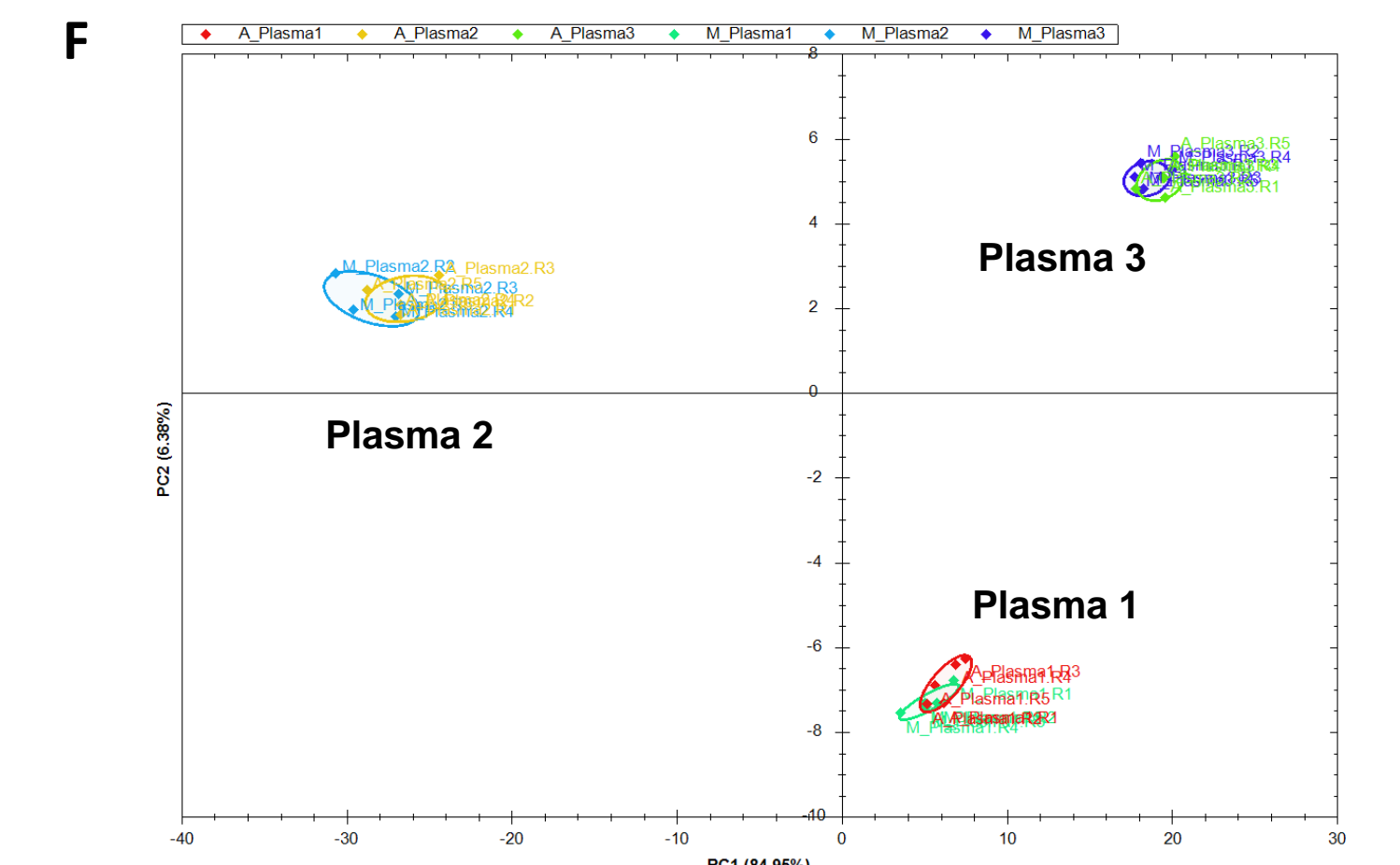
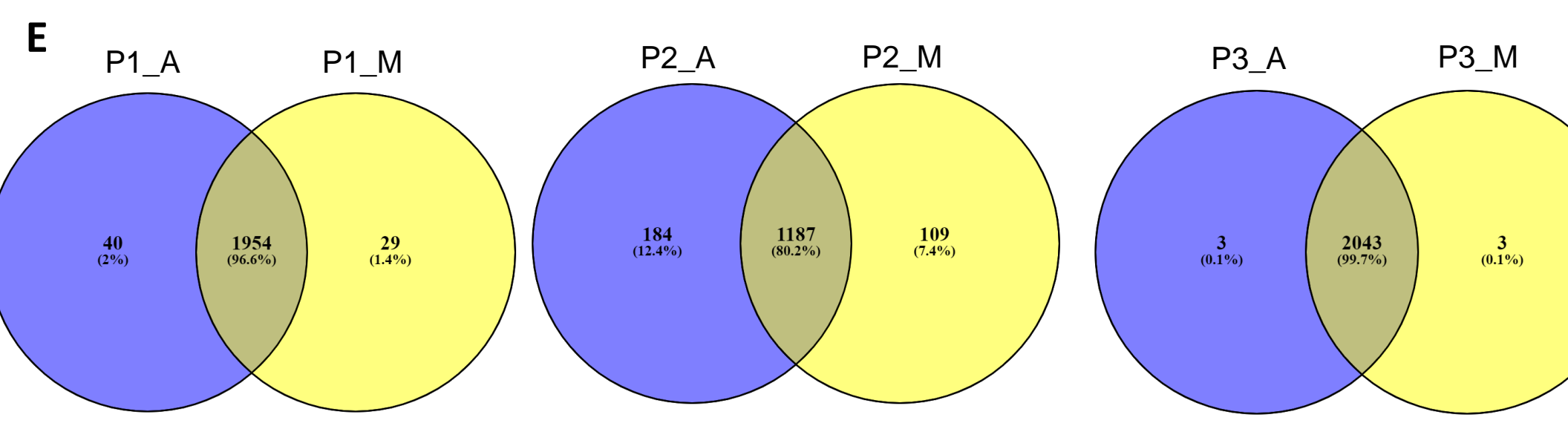
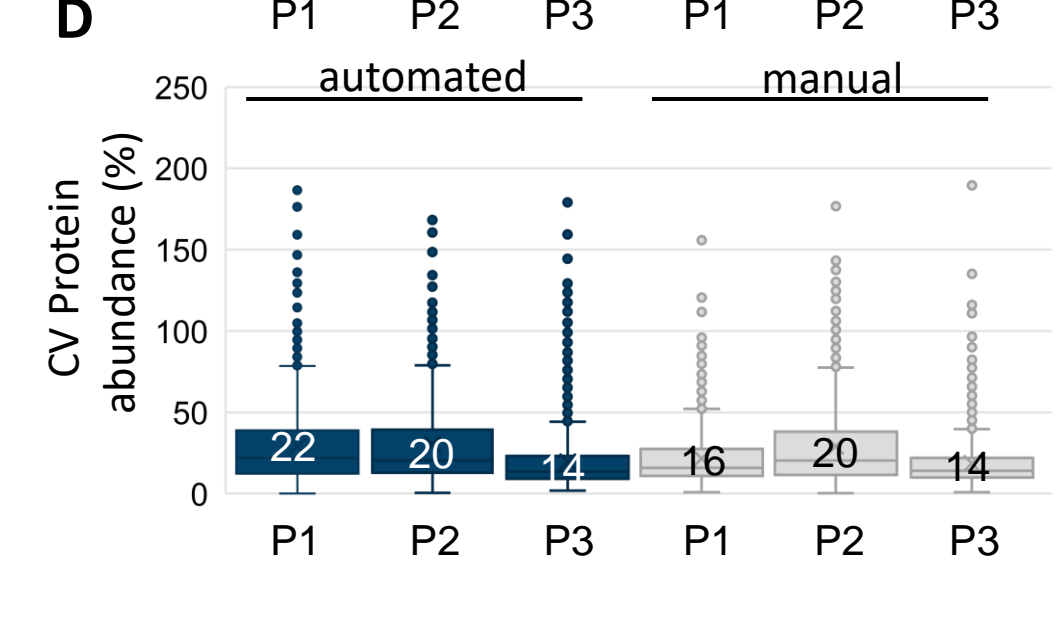
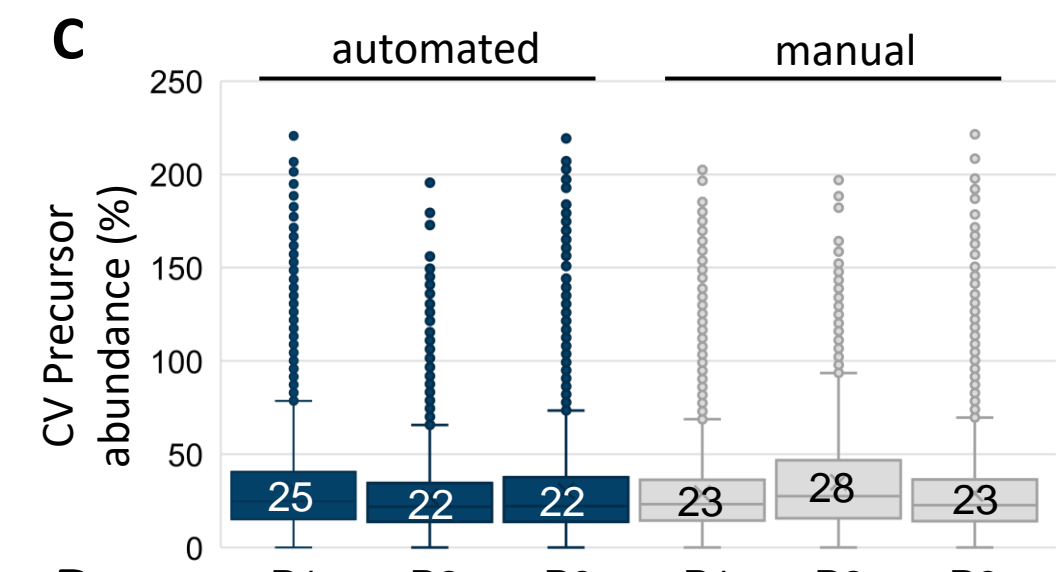
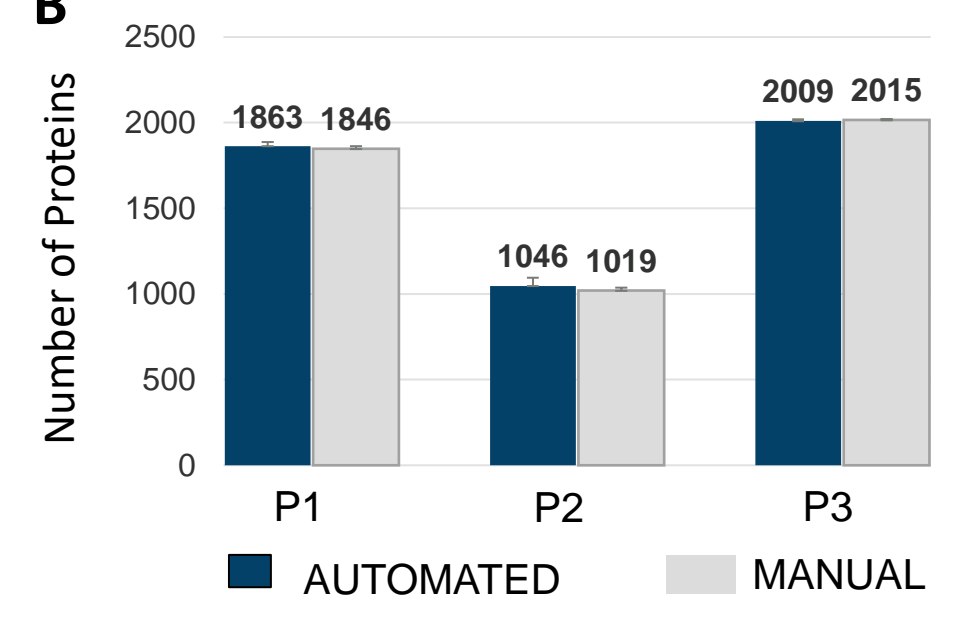
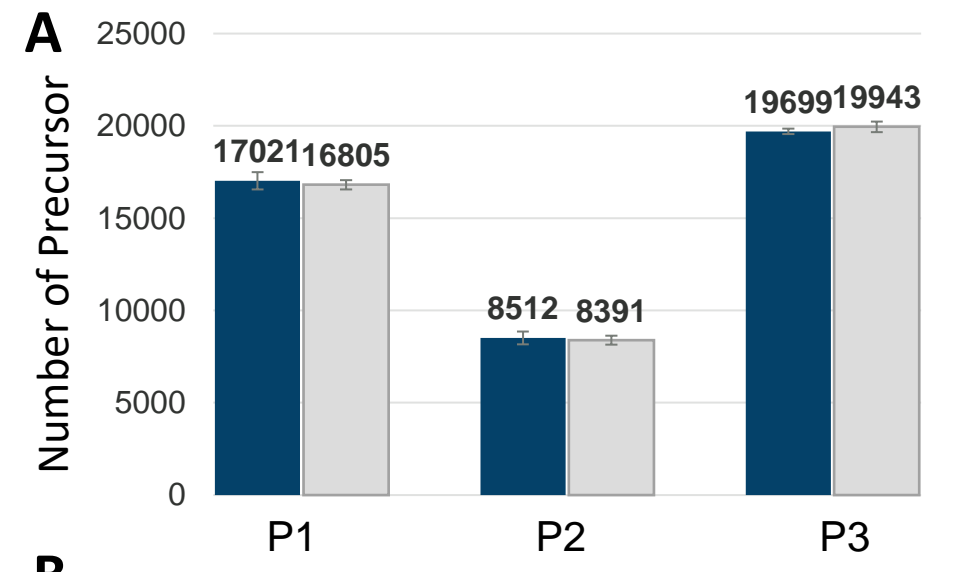


- A. Positive Pressure Module
- B. PreOmics SPE PLATE
- C. Adapter for 96-well plate
- D. Waste guide
- E. Dispenser head
- F. Collection plate

Automated peptide-clean up on the Resolvex® A200. Zoom onto workdeck, depicting positive pressure-based liquid displacement (left) and 8-channel guided dispensing of up to 11 buffers (right). Integrated version of A200 enables full end-to-end automation.

Automated ENRICH-iST workflow demonstrates excellent quantitative plasma protein characterization with on par performance to its manual counterpart.

A: Precursor identifications. > 19000 precursors have been observed for plasma samples **B: Protein identifications.** > 2000 protein groups were observed. **C: Coefficient of variation (CV, %)** of the precursor abundance. **D: CV (%)** of the protein abundance. **E: Overlap** of identified protein groups between (i) Plasma 1 automated vs Plasma 1 manual is > 96 % (ii) Plasma 2 automated vs Plasma 2 manual is > 80 % (iii) and Plasma 3 automated vs Plasma 3 manual is > 99 %. **F: Principal component analysis** reduces sample space to three clusters, driven by the plasma-origin. Moreover, automated and manual samples cluster per donor together. Thus, the manual and automated workflow preserve the same biological variability in terms of protein abundances. **G: Hierarchical clustering** depicted in heatmap shows high consistency of identifications within the same plasma sample.



Materials & Methods
Sample collection: Plasma samples were collected using EDTA from 3 healthy donors. All samples were provided in 60 µl as quintuplicates in order to evaluate the automation method reproducibility.
Sample preparation: Per replicate, 20 µl plasma was processed both manually and with the automation platform following the ENRICH-iST kit 96x HT (PreOmics) instructions. The automated was developed using the Tecan Fluent liquid handling platform in combination with Tecan's positive pressure module Resolvex® A200. The automated protocol covers the protein enrichment, the iST-based protein denaturation and digestion, together with peptide clean-up for a total of 96 samples/run within 5 hours.
LC-MS/MS analysis: Peptide samples were separated on a 10 cm column (10 cm x 75 µm, 1.5 µm, PepSep) on the nanoElute 2 nanoflow liquid chromatography system (Bruker) coupled to the Bruker's timsTOF HT via a CaptiveSpray ionization source. The sample amount injected was 400 ng in 1 µl of LC-LOAD and the separation was performed using a 20-min ACN gradient. For the dia-PASEF acquisitions, a window placement scheme consisting of 8 TIMS ramps with 3 mass ranges per ramp spanning from 400 – 1000 m/z and from a mobility range of 0.64 – 1.37 1/K0 with a cycle time of 0.5 seconds, including one MS1 frame was applied.
Data analysis: All dia-PASEF data were elaborated using Spectronaut® (v17, Biognosys®) selecting the library-free directDIA+ workflow. Factory's default settings were applied the spectra were searched against Uniprot human reviewed protein sequence database (April. 2022).

Interest?
Questions?
Please contact us!

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Mass spectrometry: Claudia.Martelli@bruker.com
Automation / Liquid Handling: Fabian.Wendt@tecan.com

The automated platform brings simplicity and profound insights to the challenging analysis of plasma empowering to find the underlying biology.

- ENRICH technology for efficient dynamic range compression coupled to robust iST-BCT sample preparation provides improved protein identification and quantification with excellent technical variability.
- High-throughput sample preparation (5 h for 96 samples) with short-gradient MS-based analysis (~ 45 samples / day) allows to scale for large cohorts.
- Manual and automation workflow show equivalent performance in terms of protein recovery, precursors/proteins ids, quantification reproducibility.

Future work:

- Automation of peptide concentration measurements & their normalization reduces sample loading time.
- Full package automating PreOmics' application portfolio (iST-BCT, iST-PSI, SP3-iST and ENRICH-iST) will be available soon.